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Research article

Prevalence of *Cryptosporidium* and *Giardia* species infections among children and calves in Chattogram, Bangladesh

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ABSTRACT

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Cryptosporidiosis and giardiasis are parasitic diseases that may significantly affect human and animal populations, notably cattle and goats. The zoonotic potential and modes of transmission of these diseases have been the subject of several epidemiological studies. We collected a total of 437 fecal specimens to determine the prevalence of cryptosporidiosis and giardiasis among human and animal populations in Bangladesh. Hence, 200 fecal samples from symptomatic children and 237 samples from calves were gathered from healthcare facilities and cattle farms in the Chattogram metropolitan area. To identify Cryptosporidium oocysts and Giardia cysts, we employed a modified Ziehl-Neelsen acidfast staining protocol (Z-N stain) for Cryptosporidium and Trichrome staining for Giardia, followed by a polymerase chain reaction (PCR) with partial amplification of SSU and tpi gene, respectively. Based on the findings of the modified Z-N stain, the prevalence of Cryptosporidium infection was determined to be 13.5% among hospitalized children with diarrhea, whereas infected calves exhibited the prevalence of 23.63%. However, the SSU gene-based polymerase chain reaction (PCR) method revealed that the frequency of Cryptosporidium infection was 9.5% among hospitalized children with diarrhea and 19.41% among infected calves. Furthermore, Trichrome staining techniques indicated that the occurrence of Giardia in children and calves was 9.5% and 19.41%, respectively. The results obtained from tpi gene-based PCR analysis revealed a prevalence rate of 9% for Giardia infection in children and 10.55% in calves. The findings from this study can be used as baseline for other researchers to perform extensive research on cryptosporidiosis and giardiasis in both human and animal populations.

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1. INTRODUCTION

Cryptosporidiosis and giardiasis are parasitic diseases that may substantially impact humans and animals, particularly cattle and goats (Ryan and Zahedi, 2019). These parasites are important waterborne pathogens due to their high prevalence rate, positive correlation with waterborne outbreaks, and their resistant pattern to multiple disinfectants (Savioli et al., 2006).

The occurrence of diarrhea in Bangladesh has been associated with the existence of both Cryptosporidium and Giardia .These diseases are often caused by two distinct parasites, Cryptosporidium and Giardia, and have been attributed to sporadic and outbreak episodes of diarrhea and malnutrition (Šlapeta, 2013). However, several epidemiological studies have investigated Cryptosporidium and Giardia transmission routes and zoonotic potential (Robertson, 2009). There are different species and genotypes of Cryptosporidium and Giardia that may infect humans, while cattle are considered to be the primary source of zoonotic transmission of these parasites (Ryan et al., 2021; Kiflevohannes et al., 2022; Faver, 2004).

Currently, there are more than 26 recognized species of Cryptosporidium, with more than 40 distinct genotypes that have not been classified as species yet (Ryan et al., 2014). Over 15 of *Cryptosporidium* have been species associated with human cryptosporidiosis, although C. hominis and C. parvum account for most cases worldwide. C hominis was shown to be the most common species responsible for diarrhea in children in research conducted in Bangladesh, Peru, Brazil, and India (Hira et al., 2011; Mbae et al., 2015; Xiao et al., 2001). A comprehensive analysis of gp60 sequencing data regarding Cryptosporidium, especially in developing countries, suggested that C. hominis accounts for around (70-90%) of human infections (Thompson, 2008; Xiao and Fayer, 2008).

In Bangladesh, Cryptosporidium and Giardia have been linked to diarrhea (Rahman et al., 1990). In cases of giardiasis, diarrhea was primarily caused by G. duodenalis assemblage A (Haque et al., 2005; Perti et al., 2009). Cryptosporidium hominis and C. parvum were found in patients with diarrhea (Perti et al., 2009). All of the studies mentioned above took place in Dhaka, one of the world's fastest expanding megacities. The presence of overcrowded slums, as well as the pollution of nearby waterways with untreated wastewater (Rahman and Hossain, 2008), may enhance (waterborne) human to human transmission. Giardia infections in rural Bangladesh have also been linked to childhood diarrhea (Hasan et al.,

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2006). An earlier study discovered a significant frequency of *Cryptosporidium* and *Giardia* in cattle. Young calves are thought to be a reservoir for these parasites, and transmission of *Cryptosporidium* and *Giardia* from cattle to cattle handlers has been reported in Bangladesh and India (Khan et al., 2011).

Previous study revealed Giardia lamblia in 68% of toddlers aged 2 to 8 months in Bangladesh (Haque et al., 2003) and identified 11.08% Giardia lamblia infection in a 2-5 year old age group in Mirpur, an urban slum district in Dhaka. Several investigations in Bangladesh have been conducted to estimate the prevalence of Giardia lamblia by direct microscopic examination (Haque et al., 2003; Alam et al., 2011). Few studies have used current diagnostic procedures such as immunofluorescence assay, enzyme-linked immunosorbent assay (ELISA), or polymerase chain reaction (PCR). A sensitive and specific diagnostic approach is required for screening Cryptosporidia and Giardia. PCR provides higher sensitivity and specificity than traditional diagnostic procedures such as direct microscopic examination) (McGlade et al., 2003).

The current study aimed to elucidate the prevalence of cryptosporidiosis and giardiasis in both the human and animal population of Chattogram by utilizing staining and polymerase chain reaction (PCR) approaches. The study also aimed to evaluate the association of different variables with the presence of water borne cryptosporidiosis and giardiasis in human and animal populations.

2. MATERIALS AND METHODS

Study area

This study was conducted on a total of 437 (n = 437) fecal samples collected from both humans and cattle suffering from diarrhea. A total of 200 (n = 200) human fecal samples were collected from the child patients (1 month to <5 years of age) with gastrointestinal discomfort, such as diarrhea, dehydration, abdominal pain, nausea, and vomiting, referring to the Chattogram Medical College Hospital, Chattogram. In the meantime, 237 (n = 237) fecal samples were also collected from calves (1 to <6 months of age) originating from different Chattogram

metropolitan area's local cattle farms, all having diarrheagenic symptoms.

Study group

The specific age categories for studying Cryptosporidium and Giardia in children and calves are designed to account for developmental and behavioral differences that influence infection risk and prevalence. For children, (1-10 months) infants and very young children, (11-15 months) children begin to explore more actively, (16-35 months) toddlers, (> 36 months) older children. The age categories for calves (1 to 15 days) newborn and very young calves, (16 to 30 days) as calves grow their immune system starts to develop, (31 to 60 days) at this stage calves are transitioning to a more solid diet, (>60 days) older calves are more robust .The ages were determined by questioning the patient's guardian and farmers.

Demographic data and sample collection

Samples were obtained from the rectum of calves using gloved fingers and were placed in sterile containers with screw caps. A stool specimen weighing between five and ten grams collected. Preventive was measures were implemented to minimize the risk of crosscontamination between specimens. To avoid direct contact with feces and any potential contaminants, gloves were used when doing this. To protect from contamination with clothing, lab coats or disposable gowns, and wear masks were used if there is in case of a possibility of splashing. We collected samples using sterile equipment and containers to prevent cross-contamination. Used clean. disposable containers made exclusively for stool collecting. Ensure that they were properly sealed to prevent leakage. The samples were promptly moved to a container with an ice bag and kept at a temperature of -20°C. In this cross-sectional study, during the period from February 2019 to June 2022, a standard pre-tested questionnaire was used to collect data on different demographic and epidemiological variables such as age, gender, clinical manifestations, symptoms, and date of specimen collection were documented.

Identification by microscopy

Before molecular testing, feces were examined

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microscopically for *Cryptosporidium* oocysts and *Giardia* cysts. All samples were stained with modified Ziehl-Neelsen acid-fast (Modified Z-N stain) (Putt, 1951) for *Cryptosporidia* and Trichrome stain (Siwila, 2017) for *Giardia* to detect the presence of *Cryptosporidium* oocysts and *Giardia* cysts, respectively. The microscopic slides were then examined with a Leica DM750 Binocular (Wetzlar, Germany) to confirm their presence. The cysts and oocysts of parasites were identified using the microscope with a magnification of 10x and 40x.

DNA extraction and PCR analysis

Genomic DNA from the whole stool samples were extracted by PureLinkTM Microbiome DNA Purification Kit (Catalog Number A29790) following the manufacturer's instruction. The extracted DNA samples were then subjected to nested polymerase chain reaction (PCR) analysis for molecular-based identification. The PCR-based identification of Cryptosporidium and Giardia was carried out by a portion of the SSU gene (240 bp) (Nolan et al., 2013) and the *tpi* gene (~530) (Sulaiman et al., 2003), respectively. Therefore, the primary amplification of SSU was performed by using 5'-GGAAGGGprimers XF2 (forward: TTGTATTTATTAGATAAAG-3') and XR2 (reverse: 5'-AAGGAGTAAGGAACAACCTC-CA-3') (Koehler et al., 2016), followed by nested amplification of SSU using the internal primers pSSUf (forward: 5'-AAAGCTCGT-AGTTGGATTTCTGTT-3') and pSSUr (reverse: 5'-ACCTCTGACTGTTAAATACRA-ATG C-3') (Nolan et al., 2010). For the primary amplification, a cycling protocol of 94°C for 5min (initial denaturation), followed by 30 cycles of 94°C for 45 s (denaturation), 45°C for 2 min (annealing), and 72°C for 1.5min (extension), with a final extension of 72°C for 10 min was employed. At the same time, the secondary amplification was achieved by employing a cycling protocol of 94°C for 5 min. followed by 35 cycles of 94°C for 30 s, 55°C for 30 s, and 72°C for 30 s, with a final extension of 72°C for 10 min (Koehler et al., 2016).

The *tpi* locus was amplified using primers AL3543 (forward: 5'-AAATTATGCCTGCTC-GTCG-3') and AL3546 (reverse: 5'-CAAACCT-TTTCCGCAAACC-3'), followed by a nested

amplification of tpi employing primers AL3544 (forward: 5'-CCCTTCATCGGTGGTAACTT-3') and AL3545 (reverse: 5'-GTGGCCACCAC-TCCCGTGCC-3') (Sulaiman et al., 2003). For the primary amplification, the cycling protocol was 94°C for 5 min (initial denaturation), followed by 30 cycles of 94°C for 45 s (denaturation), 50°C for 45 s (annealing), and 72°C for 1 min (extension) and a final extension of 72°C for 10 min. The secondary amplification of *tpi* was achieved employing 94°C for 5 min, followed by 30 cycles of 94°C for 45s, 55°C for 30 s, and 72°C for 1 min, with a final extension at 72°C for 10 min (Koehler et al., 2016).

Statistical analysis

Descriptive and analytic statistics were performed in STATA (2013) after the data was input in Microsoft Excel 2019. The Chi-square test was used to assess and compare *Cryptosporidium* and *Giardia* infection rates with different variables in both children and calves. The graphical demonstration of the data was carried out by Microsoft Excel 2019. Values of p<0.05 were considered statistically significant.

Ethics statement

This study was reviewed and approved by the Chattogram Veterinary and Animal Sciences BJVAS, Vol. 12, No. 1, January – June 2024

University Ethics Committee. The CVASU Ethics Committee approval number for the project is Memo no.-CVASU/Dir(R&E) EC/2019/39(2/10). Written informed consent was obtained from the owners for the participation of both children and animals in this study.

3. RESULTS

A total of 437 fecal specimens were examined to determine the prevalence of Cryptosporidium and Giardia infection within human and animal populations in Bangladesh. Therefore, 200 human fecal samples of symptomatic children and 237 calve samples were collected from local hospitals and farms of Chattogram respectively. According to the modified Z-N stain, the prevalence of Cryptosporidium infection among hospitalized diarrheic children and infected calves was 13.5% and 23.63%, respectively (Figure 1). However, in SSU gene-based PCR, the prevalence of Cryptosporidium hominis infection among hospitalized diarrheic children and infected calves was 9.5% and 19.41%, respectively (Figure 1). Additionally, Trichome staining revealed that the prevalence of Giardia in children and calves was 9.5% and 19.41%, respectively. While tpi gene-based PCR showed a 9% Giardia intestinalis infection prevalence in children and 10.55% in calves (Figure 1).

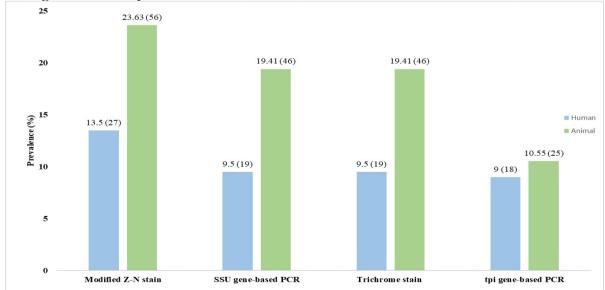


Figure 1. The prevalence of cryptosporidiosis and giardiasis in human and animal populations.

Table 1 summaries the descriptive statistics and associations between various characteristics and occurrences of cryptosporidiosis and giardiasis in the human population, as identified by the PCR method. In humans, gender-based distributions for cryptosporidiosis and giardiasis indicated that males 14.29% in cryptosporidiosis and 13.33% in giardiasis) were at a higher risk for infection than females (10.53% in cryptosporidiosis and 12.63% in giardiasis), without achieving statistical significance (*p*>0.005). Cryptosporidiosis was more prevalent in children aged 11-15 months, whereas giardiasis was more frequent in children more than >35 months. The variation of seasons significantly influences the prevalence and incidence of cryptos-poridiosis and giardiasis in the human population. Giardiasis 15.49% was shown to be more likely to spread during the winter months, while cryptosporidiosis 14.81% was more predominant in summer. However, no statistically significant differences (p>0.005) were found across the seasons. In all scenarios, children residing in

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rural areas were more susceptible to infection (14.58% in cryptosporidiosis and 13.54% in giardiasis) than their counterparts residing in areas. including citv and slum urban environments. Additionally, children who deprived with proper sanitation were more susceptible to both kind of infections (17.07% in cryptosporidiosis and 15.85% in giardiasis). Water sources were also impacted on both kind of infections, where children who consumed pond water rather than supplied and tube well water, were more prone to cryptosporidiosis 14.29% and giardiasis 42.86%. Drinking pond water is often associated with higher risks of waterborne diseases, including those caused by protozoa like Cryptosporidium and Giardia, due to potential contamination with pathogens. Nevertheless, no statistically significant (p>0.05) associations were identified in both circumstances. Children with a history of being mix (breastfed and formula) are more susceptible to both types of infections, where in case of giardiasis significant differences were recorded (p>0.05).

Table 1. Descriptive statistics and association of different variables with cryptosporidiosis and giardiasis in the human population diagnosed by PCR

Variable	Level	Total	No. positive for	p-value	No. positive	p-value
		observation	Cryptosporidium	for χ ²	for <i>Giardia</i>	for χ^2
		(%)	(%)	test	(%)	test
Gender	Female	95 (47.50)	10 (10.53)	0.42	12 (12.63)	0.88
	Male	105 (52.50)	15 (14.29)	-	14 (13.33)	
Age (months)	1 to 10	55 (27.50)	7 (12.73)	0.24	6 (10.19)	0.85
	11 to 15	45 (22.50)	9 (20.00)	-	5 (11.11)	
	16 to 35	50 (25.00)	6 (12.00)	_	7 (14.00)	
	>36	50 (25.00)	3 (6.00)	-	8 (16.00)	
Season of sample collection	Summer	54 (27.55)	8 (14.81)	0.59	3 (5.56)	0.06
	Winter	142 (72.45)	17 (11.97)	_	22 (15.49)	-
Residence	City	76 (38.00)	9 (11.84)	0.56	10 (13.16)	0.92
	Slum	28 (14.00)	2 (7.14)	_	3 (10.71)	
	Village	96 (48.00)	14 (14.58)	-	13 (13.54)	
Source of water	Pond	8 (4.00)	1 (14.29)	0.73	3 (42.86)	0.06
	Supply	98 (49.25)	10 (10.20)	_	11 (11.22)	
	Tube well	94 (47.24)	13 (13.83)	-	12 (12.77)	-
History of breastfeeding	Yes	115 (57.50)	14 (12.17)	0.55	14 (12.17)	0.02
	No	82 (41.00)	10 (12.20)	-	10 (12.20)	
	Mix	3 (1.50)	1 (33.33)	_	2 (66.67)	-
History of pet	Yes	96 (48.00)	14 (14.58)	0.39	10 (10.42)	0.30
·	No	104 (52.00)	11 (10.58)	-	16 (15.38)	-
History of	Yes	57 (28.50)	5 (8.77)	0.31	5 (8.77)	0.26
anthelmintics	No	143 (71.50)	20 (13.99)	-	21 (14.69)	-

The correlations between clinical characteristics and disease prevalence were also investigated and analyzed. The condition was not shown to be associated with a history of nausea, anorexia. abdominal pain, vomiting. or dehydration (Table 2). In case of calves the frequency of cryptosporidiosis (13.19%) and giardiasis (2.78%) was reported higher in females in comparison to males (10.75% and 2.15% respectively) although the association had no significant differences (p>0.05). The frequency of cryptosporidiosis was found to be relatively high in calves aged between 31 and 60 days; and in case of Giardia it was higher in calves aged between 16-31 days, though the association was not significant across the age groups (p>0.05). At the farm level, the frequency of cryptosporidiosis was shown to be independent of many factors, including the education level of owners, topography, types of calf housing, and floor type (p>0.05) (Table 4). Nevertheless, a substantial prevalence of cryptosporidiosis was seen in calves residing in hilly regions (17.65%) and giardiasis was seen 5.88%. Furthermore, it was shown that the incidence of cryptosporidiosis was comparaBJVAS, Vol. 12, No. 1, January – June 2024

tively elevated in calves that were exposed to groundwater 15.87% as opposed to pond 0.00% or supply water 11.54% and also in giardiasis it was high in ground water sources. However, no statistically significant difference was evident between these variables (p>0.05).

A farm's drainage system appeared not to have any effect on cryptosporidiosis and giardiasis prevalence. Notably, calves that had fortnightly bedding changes had a higher frequency (17.39 %) of cryptosporidiosis but not in giardiasis (2.17%) this difference was not statistically although significant (p>0.05). There was also an increase in the frequency (28%) of cryptosporidiosis among calves that did not share feeding utensils but not in giardiasis. Moreover, the frequency of cryptosporidiosis was (66.67%) higher in calves that were hand-fed compared to suckling-fed (13.56%). And whereas in case for gardiasis the prevalence was relatively much (33.33%) higher in hand fed calves compared to suckler calves (2.82%). However in case of giardiasis there were significant differences (p>0.05) in comparison to between the two feeding methods (Table 4).

Clinical findings	Level	Total observation (%)	No. positive for Cryptosporidium (%)	<i>p-value</i> for χ ² test	No. positive for <i>Giardia</i> (%)	<i>p-value</i> for χ^2 test
History of nausea	Yes	68 (34)	6 (8.82)	0.26	8 (11.76)	0.71
	No	132 (66)	19 (14.39)	_	18 (13.64)	_
Abdominal discomfort	Yes	76 (38.00)	10 (13.16)	0.82	7 (9.21)	0.21
	No	124 (62.00)	15 (12.10)	_	19 (15.32)	_
Anorexia	Yes	15 (7.50)	2 (13.33)	0.91	2 (13.33)	0.97
	No	185 (92.50)	23 (12.43)	_	24 (12.97)	_
Vomiting	Yes	68 (34.0)	13 (19.12)	0.04	11 (16.18)	0.34
	No	132 (66.0)	12 (9.09)	_	15 (11.36)	_
Dehydration	Yes	103 (51.50)	15 (14.56)	0.36	15 (14.56)	0.50
	No	97 (48.50)	10 (10.31)	_	11 (11.34)	_

Table 2. Description of clinical findings in cryptosporidiosis and giardiasis in the human population diagnosed by PCR

4. DISCUSSION

Cryptosporidium and *giardia* have the potential to cause clinical disease in calves and to be transmitted to other animal species and humans. Detection of *Cryptosporidium* and *Giardia* in children and calves therefore may be of great public health significance, as humans and animals share sometimes the same premises in rural settings. Based on the findings of the

modified Z-N stain, the prevalence of *Cryptosporidium* infection was determined to be 13.5% among hospitalized children, whereas infected calves exhibited the prevalence of 23.63%. However, the *SSU* gene-based PCR method revealed that the frequency of *Cryptosporidium* infection was 9.5% among hospitalized children with diarrhea and 19.41% among infected calves.

Variable	Level	Total observation (%)	No. positive for Cryptosporidium (%)	<i>p-value</i> for χ ² test	No. positive for <i>Giardia</i> (%)	<i>p-value</i> for χ ² test
Sex	Female	144 (60.76)	19 (13.19)	0.58	4 (2.78)	
	Male	93 (39.24)	10 (10.75)		2 (2.15)	0.77
Age	1 to 15	49 (20.68)	6 (12.24)	0.35	1 (2.04))	
(days)	16 to 30	58 (24.47)	7 (12.07)		3 (5.17)	
	31 to 60	70 (29.54)	12 (17.14)		0	0.30
	>60	60 (25.32)	4 (6.67)		2 (3.33)	

Table 3. Descriptive statistics and association of different variables with *Cryptosporidium* and *Giardia* at the animal level diagnosed by PCR

This study showed a prevalence of 13.5% for Cryptosporidium in children with persistent diarrhea diagnosed by the modified Ziehl-Neelsen staining method, where this percentage agrees with the results of (Tairsh et al., 2017) who reported the prevalence of 12.85%. However, the previous study by Ehsan et al. (2015) suggested that the prevalence of the SSU gene was found to be 5% each in calves infected with Cryptosporidium infection. While the prevalence of 1-4% was reported in Europe and North America, and it was 3-20% in Australia, Asia, Africa, and Central and South America (WHO, 2009). Another study (Morgan et al., 1998) reported 36% by microscopic and 29% by PCR method. Furthermore, Trichrome staining techniques indicated that the occurrence of Giardia in children and calves was 9.5% and 19.41%, respectively. The findings are similar to the previous study reported by Datta (2024). In contrast the previous study by Ehsan et al. (2015) reported that the prevalence of the tpi gene was found to be 21.7% in calves infected with Giardia infection. In this study, tpi genebased PCR analysis revealed a prevalence rate of 9% for Giardia infection in children and 10.55% in calves hence the results were in agreement with (Alseady et al., 2023). Usually the PCR is substantially more sensitive than staining methods. However, the outcomes of the study for cryptosporidiosis and giardiasis staining results were higher than the PCR result.

This difference may be due to sample size and diagnostic techniques because PCR may yield less accurate results than microscopy due to issues with sample preparation or PCR inhibitors, microscopy can directly visualize parasites regardless of these conditions.

Furthermore, PCR approaches may not target all parasite strains or stages, resulting in different detection results.

Cryptosporidiosis and giardiasis are gastrointestinal diseases caused by parasitic protozoa such Cryptosporidium and Giardia, respectively (Ehsan et al., 2015).Understanding the factors that influence the prevalence of these infections is essential for public health efforts. However, the present study delves into gender-based distributions, age, season, urban-rural residence, breastfeeding, water sources. and pet ownership/exposure to anthelmintics, analyzing their potential impact on the prevalence of cryptosporidiosis and giardiasis in the human population. The data suggests a slight genderbased difference in infection rates, with males showing a marginally higher risk for both cryptosporidiosis 14.29% and giardiasis 13.33% compared to females 10.53% and 12.63 %, respectively. However, these differences were not statistically significant (p>0.05). It is important to note that gender alone may not be a primary determinant of infection risk, and other factors may play a more significant role. Regarding giardiasis, the findings are similar to the previous study by (Suman et al., 2011). Additionally, research conducted in India and Nigeria has shown a higher prevalence of giardiasis infection in males than females (Dwivedi et al., 2007). The data also indicates that cryptosporidiosis is more prevalent in children aged 11-15 months, whereas giardiasis is more frequent in children older than 35 months. The prevalence of giardiasis is analogous to other previous studies by Haque et al. (2003) and Suman et al. (2011).

Variable	Level	Total observation (%)	No. positive for Cryptosporidium (%)	<i>p- value</i> for χ ² test	No. positive for <i>Giardia</i> (%)	<i>p-value</i> for χ ² test
Owners'	Illiterate	7 (3.93)	0	0.16	0	
education	Primary	24 (13.48)	3 (12.50)	-	0	
	Secondary	42 (23.60)	7 (16.67)	_	2(4.76)	
	Higher Secondary	32 (17.98)	8 (25.00)	_	1(3.13)	0.55
	Graduation	44 (24.58)	3 (6.82)	_	3(6.82)	
	Post-graduation	30 (16.85)	5 (16.67)	-	0	
Topography	Hilly	34 (18.89)	6 (17.65)	0.55	4(5.88)	0.35
	Plain land	146 (81.11)	20 (13.70)	-	2 (2.47)	
Types of calf	Closed barn	116 (64.44)	17 (14.66)	0.99	4 (3.45)	
house	Open barn	15 (8.33)	2 (13.33)	-	0	0.74
	Partial open	49 (27.22)	7 (14.29)	-	2 (4.08)	
Floor-type	Brick	92 (51.11)	13 (14.13)	0.97	3 (3.26)	
	Concrete	73 (40.56)	11 (15.07)	-	1 (1.37)	0.06
	Muddy/jute/wood	15 (8.33)	2 (13.33)	-	2 (13.33)	
Source of	Ground	126 (70.0)	20 (15.87)	0.64	4 (3.17)	
drinking	Pond or stream	2 (1.11)	0	-	0	0.94
water	Supply	52 (28.89)	6 (11.54)	-	2 (3.85)	
Type of	None	32 (17.78)	5 (15.63)	0.75	2 (6.25)	
drainage	Open	118 (65.56)	18 (15.25)	-	3 (2.54)	0.59
system	Sub-surface	30 (16.67)	3 (10.00)	-	1 (3.33)	
Frequency of bedding change	Never	18 (10.0)	2 (11.11)	0.72	1 (5.56)	
	Weekly	111 (61.67)	16 (14.41)	-	3 (2.70)	
	Fortnightly	46 (25.56)	8 (17.39)	-	1 (2.17)	0.18
	Monthly	5 (2.78)	0	-	1 (20.00)	
Hygiene of	Not shared	25 (13.89)	7 (28.0)	0.11	1 (4.00)	
calf feeding utensils	Shared and disinfected	45 (25.0)	6 (13.33)	-	0	0.35
	Shared and washed with water	110 (61.11)	13 (11.82)	-	5 (4.55)	
Feeding of	Hand feeding	3 (1.67)	2 (66.67)	0.01	1 (33.33)	
milk	Suckling	177 (98.33)	24 (13.56)	0.01	5 (2.82)	0.004*

Table 4. Descriptive statistics and association of different variables with *Cryptosporidium* and *Giardia* at farm level diagnosed by PCR

Cryptosporidiosis peaks in the 11-15 month age group possibly due to their developing immune systems and increased exposure to infection through behaviors like mouthing objects and interacting in group settings. However, the potential factors contributing to this agedependent trend are likely associated with the behaviors of children. Another potential factor contributing to the increased incidence of infections in children might be the insufficient development of efficient immunity (Suman et al., 2011). However, no statistically significant differences were found. This suggests that while age may contribute to susceptibility, it is not a sole determinant of infection risk. Seasonal variation substantially impacts the prevalence incidence of cryptosporidiosis and and giardiasis, where the summer season was found to be more feasible for Cryptosporidium and winter for Giardia infections. Giardiasis is more prevalent in the winter possibly because milder temperatures preserve Giardia cysts in the environment and increase indoor activity, whereas cryptosporidiosis is more common in the summer because warmer temperatures and increased recreational water use encourage the survival and spread of *Cryptosporidium* oocysts. Children residing in rural areas appear to be

more susceptible to infection (14.58% in cryptosporidiosis and 13.54% in giardiasis) than their urban counterparts. However, the lack of statistical significance (p>0.05) suggests that while residence may play a role, other factors are at play, too, including access to healthcare and sanitation facilities. Although the lowest prevalence of both the protozoa among the slum ideal factors dwellers usually for the development but in some areas, targeted public health interventions may have improved water quality, sanitation, and hygiene practices, even in slum environments. These improvements can significantly reduce the prevalence of protozoan infections and also residents in slums may develop partial immunity due to repeated lowlevel exposures could lead to lower reported prevalence rates.

Notably, children lack of proper sanitation facilities was more susceptible to both kind infections. Cryptosporidium and Giardia are often spread by contaminated water; hence, it was interesting to see whether the water source affected the likelihood of infection. In this study, no statistically association was seen in the water source, though, children who had consumed pond water, were more prone to cryptosporidiosis and giardiasis. Two other studies conducted in urban and semi-urban regions in Brazil (Pereira et al., 2002) and Guinea Bissau (Mølbak et al., 1994) have also similarly shown no significant correlation between cryptosporidiosis and either the source or type of water supply. Children who had previously been nursed and formula fed were shown to be more susceptible to both types of illnesses, but there were significant differences (p>0.05) in giardiasis. Children with a history of mix feeding have a relatively high prevalence of Cryptosporidium and Giardia infection which could be attributed to supplemental feeding and difficulty with exclusive breastfeeding.

According to this study, cryptosporidiosis was more prevalent in calves than children. The present study also showed that cryptosporidiosis was expected in 31–60-day-old calves, which is supported by other previous studies by (Gow and Waldner, 2006; Khair et al., 2014; Maldonado-Camargo et al., 1998; Paul et al., 2009); and (Swai and Schoonman, 2010). The increased incidence of *cryptosporidium* and

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giardia in calves aged 16 to 30 days could be attributed to increased exposure to contaminated environment when their immune systems are still maturing. This age range frequently coincides with the period during which calves are more active and contact with polluted feed or water sources. As a fore mentioned, the prevalence of cryptosporidiosis in females was high (13.19%), which is opposite to the study conducted by (Nouri and Toroghi, 1991), it was shown that male diarrheic calves had a greater incidence of infection compared to female calves (Nouri and Toroghi, 1991). However, the prevalence of cryptosporidiosis did not exhibit a statistically significant difference based on the gender of calves, which is consistent with findings reported by previous studies of (Khair et al., 2014; Rahaman et al., 1984).

5. CONCLUSION

The study aims to inform public health strategies and improve infection control measures for both humans and animals. Examining diarrhoeic samples from both children and calves in the same city for Cryptosporidium and Giardia aims to highlight the zoonotic importance of these parasites. By investigating infections in both human and animal populations, the study seeks to demonstrate how these parasites can be transmitted between species and affect both groups. This approach underscores the significance of zoonotic transmission and helps in understanding the broader implications for animal management. public health and Regarding this study, the epidemiology of cryptosporidiosis and giardiasis in Bangladesh indicates a complex, challenging context. epidemiology of Therefore, the cryptosporidiosis and giardiasis in Bangladesh highlights the urgent need for extensive public health efforts to combat these enduring and disabling parasitic illnesses. The well-being and contentment of the people will improve, and the entire country will move closer to its long-term aim of sustainable development if this objective is met.

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REFERENCES

- Alam, M. M., Ilias, M., Siddique, M. A., Kabir, M. M., Nazib, F. and Khan, M. G. M. 2011. Genotype-specific detection of Giardia lamblia in stool samples of diarrhoeal and non-diarrhoeal patients in Dhaka, Bangladesh. Dhaka University Journal of Biological Sciences, 20(2): 183-189.
- Alseady, H. H., Al-Dabbagh, S. M. and Marhash, A. D. 2023. Prevalence and molecular characterization of Giardia intestinalis isolated from children and calves in Babylon province, Iraq. Veterinary World, 16(9): 1781.
- Datta, P., Garg, P., Lal Bhasin, S., Malhotra, P., Rana, S. S. and Khurana, S. 2024. Modified trichrome stain for faster and improved detection of intestinal protozoan parasites. Tropical Doctor, 54(2): 139-146.
- Dwivedi, K. K., Prasad, G., Saini, S., Mahajan, S., Lal, S. and Baveja, U. K. 2007. Enteric opportunistic parasites among HIV infected individuals: associated risk factors and immune status. Japanese Journal of Infectious Diseases, 60(2/3): 76.
- Ehsan, A. M., Geurden, T., Casaert, S., Parvin, S. M., Islam, T. M., Ahmed, U. M., Levecke, B., Vercruysse, J. and Claerebout, E. 2015. Assessment of Zoonotic Transmission of Giardia and Cryptosporidium between Cattle and Humans in Rural Villages in Bangladesh. PLOS ONE, 10: 0118239.
- Fayer, R. 2004. Cryptosporidium: a water-borne zoonotic parasite. Veterinary Parasitology. 126: 37-56.
- Gow, S., Waldner, C. 2006. An examination of the prevalence of and risk factors for shedding of Cryptosporidium spp. and Giardia spp. in cows and calves from western Canadian cow-calf herds. Veterinary Parasitology, 137:50-61.
- Hasan, K. Z., Pathela, P., Alam, K., Podder, G., Faruque, S. M., Roy, E. and Sack, R. B. 2006. Aetiology of diarrhoea in a birth cohort of children aged 0-2 year (s) in rural Mirzapur, Bangladesh. Journal of Health, Population and Nutrition, pp. 25-35.
- Haque, R., Roy, S., Kabir, M., Stroup, S. E., Mondal, D. and Houpt, E. R. 2005. Giardia assemblage A infection and diarrhea in Bangladesh. The

BJVAS, Vol. 12, No. 1, January – June 2024

Journal of Infectious Diseases, 192(12): 2171-2173.

- Haque, R., Mondal, D., Kirkpatrick, B. D., Akther, S., Farr, B. M., Sack, R. B., and Petri, W. A. 2003. Epidemiologic and clinical characteristics of acute diarrhea with emphasis on Entamoeba histolytica infections in preschool children in an urban slum of Dhaka, Bangladesh. The American Journal of Tropical Medicine and Hygiene, 69(4): 398-405.
- Hira, K. G., Mackay, M. R., Hempstead, A. D., Ahmed, S., Karim, M. M., O'Connor, R. M., Hibberd, P. L., Calderwood, S. B., Ryan, E. T., Khan, W. A. and Ward, H. D. 2011. Genetic Diversity of Cryptosporidium spp. from Bangladeshi Children. Journal of Clinical Microbiology, 49: 2307-2310.
- Khair, A., Alam, M. M., Rahman, A. K. M. A., Shahiduzzaman, M., Parvej, M. and Chowdhury, E.H. 2014. Prevalence of Cryptosporidiosis in Crossbred Calves in Two Selected Areas of Bangladesh. Bangladesh Journal of Veterinary Medicine, 12: 185-190.
- Khan, S. M., Debnath, C., Pramanik, A. K., Xiao, L., Nozaki, T. and Ganguly, S. 2011. Molecular evidence for zoonotic transmission of Giardia duodenalis among dairy farm workers in West Bengal, India. Veterinary Parasitology, 178(3-4): 342-345.
- Kifleyohannes, T., Nødtvedt, A., Debenham, J. J., Tysnes, K. R., Terefe, G. and Robertson, L. J. 2022. Cryptosporidium and Giardia infections in humans in Tigray, Northern Ethiopia: an unexpectedly low occurrence of anthropozoonotic transmission. Acta Tropica, 231: 106450.
- Koehler, A. V., Haydon, S. R., Jex, A. R. and Gasser, R. B. 2016. Cryptosporidium and Giardia taxa in faecal samples from animals in catchments supplying the city of Melbourne with drinking water (2011 to 2015). Parasites & Vectors, 9: 315.
- Letunic, I. and Bork, P. 2019. Interactive Tree Of Life (iTOL) v4: recent updates and new developments. Nucleic Acids Research, 47(W1): W256-W259.
- Maldonado-Camargo, S., Atwill, E. R., Saltijeral-Oaxaca, J. A. and Herrera-Alonso, L. C. 1998. Prevalence of and risk factors for shedding of Cryptosporidium parvum in Holstein Freisian dairy calves in central México. Preventive Veterinary Medicine, 36: 95-107.

- Mbae, C., Mulinge, E., Waruru, A., Ngugi, B., Wainaina, J. and Kariuki, S. 2015. Genetic Diversity of Cryptosporidium in Children in an Urban Informal Settlement of Nairobi, Kenya. PLOS ONE, 10: 0142055.
- McGlade, T. R., Robertson, I. D., Elliot, A. D. and Thompson, R. C. A. 2003. High prevalence of Giardia detected in cats by PCR. Veterinary Parasitology, 110(3-4): 197-205.
- Mølbak, K., Aaby, P., Højlyng, N. and da Silva, A. P. 1994. Risk factors for Cryptosporidium diarrhea in early childhood: a case-control study from Guinea-Bissau, West Africa. American Journal of Epidemiology, 139: 734-740.
- Morgan, F. U., Pallant, L., Dwyer, B. W., Forbes, D. A., Rich, G. and Thompson, R. C. A. 1998. Comparison of PCR and microscopy for detection of Cryptosporidium parvum in human fecal specimens: clinical trial. Journal of Clinical Microbiology, 36(4): 995-998.
- Nolan, M. J., Jex, A. R., Koehler, A. V., Haydon, S. R., Stevens, M. A. and Gasser, R. B. 2013.
 Molecular-based investigation of *Cryptosporidium* and *Giardia* from animals in water catchments in southeastern Australia. Water Research, 47: 1726-1740.
- Nolan, M. J., Jex, A. R., Pangasa, A., Young, N. D., Campbell, A. J., Stevens, M. and Gasser, R. B. 2010. Analysis of nucleotide variation within the triose-phosphate isomerase gene of Giardia duodenalis from sheep and its zoonotic implications. Electrophoresis, 31: 287-298.
- Nouri, M. and Toroghi, R. 1991. Asymptomatic cryptosporidiosis in cattle and humans in Iran. The Veterinary Record, 128: 358-359.
- Paul, S., Chandra, D., Tewari, A., Banerjee, P.s., Ray, D., Raina, O. and Rao, J. 2009. Prevalence of Cryptosporidium andersoni: A molecular epidemiological survey among cattle in India. Veterinary Parasitology, 161: 31-35.
- Pereira, M. D., Atwill, E. R., Barbosa, A. P., Silva, S.A. and García-Zapata, M.T. 2002. Intrafamilial and extra-familial risk factors associated with Cryptosporidium parvum infection among children hospitalized for diarrhea in Goiânia, Goiás, Brazil. The American Journal of Tropical Medicine and Hygiene, 66: 787-793.
- Petri Jr, W. A., Haque, R., Mondal, D., Karim, A., Molla, I. H., Rahim, A. and Snider, C. 2009. Prospective case-control study of the association between common enteric protozoal parasites and diarrhea in Bangladesh. Clinical Infectious Diseases, 48(9): 1191-1197.

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- Putt, F. A. 1951. A Modified Ziehl-Neelsen Method: For Demonstration of Leprosy Bacilli and Other Acid-Fast Organisms*. American Journal of Clinical Pathology, 21::92-95.
- Rahman, S. and Hossain, F. 2008. Spatial assessment of water quality in peripheral rivers of Dhaka City for optimal relocation of water intake point. Water Resources Management, 22: 377-391.
- Rahman, M., Shahid, N. S., Rahman, H., Sack, D. A., Rahman, N. and Hossain, S. 1990.
 Cryptosporidiosis: a cause of diarrhea in Bangladesh. The American Journal of Tropical Medicine and Hygiene, 42(2): 127-130.
- Rahaman, A. S., Sanyal, S. C., Al-Mahmud, K. A., Sobhan, A., Hossain, K. S. and Anderson, B. C. 1984. Cryptosporidiosis in calves and their handlers in Bangladesh. Lancet (London, England), 2: 221.
- Robertson, L. J. 2009. Giardia and Cryptosporidium infections in sheep and goats: a review of the potential for transmission to humans via environmental contamination. Epidemiology and Infection, 137: 913-921.
- Ryan, U., Xiao, L., Cacciò, S. and Widmer, G. 2014. Cryptosporidium: parasite and disease, 1st ed., Springer, Vienna, 564p.
- Ryan, U., Zahedi, A., Feng, Y. and Xiao, L. 2021. An Update on Zoonotic Cryptosporidium Species and Genotypes in Humans. Animals, 11:3307.
- Ryan, U. N. A., Fayer, R. and Xiao, L. 2014. Cryptosporidium species in humans and animals: current understanding and research needs. Parasitology, 141: 1667-1685.
- Savioli, L., Smith, H., Thompson, A., 2006. Giardia and Cryptosporidium join the 'Neglected Diseases Initiative'. Trends in Parasitology, 22: 203-208.
- Siwila, J. 2017. Giardiasis: Livestock and Companion Animals. In: Current Topics in Giardiasis, (ed.). A.J. Rodriguez-Morales, IntechOpen, United Kingdom, 70874p.
- Šlapeta, J., 2013. Cryptosporidiosis and Cryptosporidium species in animals and humans: A thirty colour rainbow? International Journal for Parasitology, 43: 957-970.
- Sulaiman, I. M., Fayer, R., Bern, C., Gilman, R. H., Trout, J. M., Schantz, P. M., Das, P., Lal, A. A. and Xiao, L. 2003. Triosephosphate isomerase gene characterization and potential zoonotic transmission of Giardia duodenalis. Emerging Infectious Diseases, 9: 1444-1452.

- Suman, M. S. H., Alam, M. M., Pun, S. B., Khair, A., Ahmed, S. and Uchida, R. Y. 2011. Prevalence of Giardia lamblia infection in children and calves in Bangladesh. Bangladesh Journal of Veterinary Medicine, 9(2): 177-182.
- Swai, E. S. and Schoonman, L. 2010. Investigation into the Prevalence of Cryptosporidium Infection in Calves among Small-Holder Dairy and Traditional Herds in Tanzania. Veterinary Medicine International 2010: 676451.
- Tairsh, H. R., Abdul, A. L., Al-A-asady, R. A., ALzeyadi, M. J. and Ali, H. 2017. Identification of Cryptosporidium spp. infections in children with persistent diarrhea by modified ziehl-neelsen stain method and pcr technique. Eur. J. Pharm. Med. Res., 4(2): 208-215.
- Thompson, R. C., Fayer, R. and Xiao L. 2008. (eds): Cryptosporidium and cryptosporidiosis. Parasites & Vectors - Parasites Vectors 1: 1

BJVAS, Vol. 12, No. 1, January – June 2024

- Website. Available: http://www.unicef.org/ bangladeshUNICEF/Arsenic.pdf. Accessed 2024 feb.14.
- WHO (World Health Organization). 2009. Risk assessment of Cryptosporidium in drinking water (No. WHO/HSE/WSH/09.04). World Health Organization.
- Xiao, L., Bern, C., Limor, J., Sulaiman, I., Roberts, J., Checkley, W., Cabrera, L., Gilman, R. H. and Lal, A. A. 2001. Identification of 5 types of Cryptosporidium parasites in children in Lima, Peru. The Journal of Infectious Diseases, 183: 492-497.
- Xiao, L. and Fayer, R. 2008. Molecular characterisation of species and genotypes of Cryptosporidium and Giardia and assessment of zoonotic transmission. Int J Parasitol., 38: 1239-1255.